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Release of offensive odorants from the combustion of barbecue charcoals

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ABSTRACT

A number of offensive odorants including volatile organic compounds (VOCs), reduced sulfur compounds (RSCs), carbonyls, and ammonia were measured along with several reference pollutants (like benzene (B), CS₂, SO₂, CO, and total hydrocarbon (THC)) from combusted fumes of barbecue charcoals produced from five different countries (Korea, China, Indonesia, Malaysia, and the US). Although the emission concentrations of most odorants were generally below the reference guideline set by the malodor prevention law in Korea, the mean concentration of some aldehydes (acetaldehyde, propionaldehyde, and isovaler-aldehyde) and ammonia exceeded those guidelines. As such, aldehydes were the most dominant odorant released from charcoal combustion followed by VOC and ammonia. If odorant levels of charcoal products are compared, there are great distinctions between the products of different countries. If comparison is made using the concept of the sum of odor intensity (SOI), the magnitude of SOI for the charcoal products from the five different countries varied in the order of 4.30 (Korea), 3.10 (Indonesia), 2.97 (China), 2.76 (Malaysia), and 2.76 (the US).

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1. Introduction

Charcoal is a conventional term for a char obtained from wood, peat, coal, or some related natural organic material through carbonization. This process allows for the conversion of organic matter to solid residues with increased elemental carbon content via pyrolysis [1]. As charcoal has high heating values with low smoke and dust relative to wood [2], it is used extensively for cooking purposes, especially in barbequing. As an integral part of leisure activities and/or ceremonies, charcoal barbecue foods are prepared for different cultural activities in many countries. In some countries (i.e., Korea, China, etc.), traditional barbecue restaurants are also very popular, wherein charcoal is used as the main fuel for barbecuing. Although charcoal is not the prime source of cooking energy in many developed countries, it remains as one of the most commonly selected fuels for cooking in the present world.

Charcoal is made up of various types of organic and inorganic compounds such as hydrocarbons, sulfur, water, and oxygen along with numerous trace elements [1]. Recent investigations of charcoals also revealed that its combustion fume can act as a potential source of volatile organic compounds (VOCs), carbonyls, trace metals (including mercury), and polycyclic aromatic hydrocarbon (PAH) [3–7]. As a result of high airborne pollutant emissions from charcoal combustion, the Canadian government listed charcoal as a hazardous material [8]. According to the Canadian regulation guidelines, the toxic fumes of charcoal may occur and cause death if the charcoal is burnt without adequate ventilation systems.

In this study, the emission characteristics of odorant pollutants were investigated from gases released from the combustion of charcoal products. To this end, the emission concentrations of key offensive odorants were measured from a total of 15 different barbecue charcoal products commercially available in the Korean market. In a number of previous studies, we investigated the emission characteristics of several pollutant groups including aromatic volatile organic compounds and carbonyls [4] and trace metals including mercury [6,7]. The metallic content in raw (unburnt) charcoal products was also investigated [5]. In an effort to extend the database of pollutant emissions from charcoal use, odorant emissions due to charcoal combustion were analyzed to unveil the basic features of this odorant nuisance in our everyday life activities.

2. Materials and methods

2.1. Sample collection

In order to assess emission concentration levels of odorant gases (Table 1) in charcoal fumes, a series of experiments were carried out by combusting 15 different charcoal products from 5 different countries (6 from Korea, 4 from China, 3 from Indonesia, 1 from Malaysia, and 1 from the US), all of which are commonly available in the S. Korean market. The collection of sample gases was made by following the procedures described in previous studies [4]. For the collection of gaseous samples, charcoal

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| Table 1 |
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| List of od |

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|---------|-----------|---------------|-----------|------------|-------------|--------------|------------|--------------|
| List of | odorous n | ollutants and | reterence | components | emitted tr | om charcoal | comples in | this study d |
| LISC U | | onutants and | reference | components | children in | onn charcoar | sampies m | uns study. |
| | | | | | | | | |

| Group | Order ^b | Full name | Short name | CAS Number | Chemical formula | Molecular weight (g mol ⁻¹) | Odor threshold (ppb) | Permissible concentration (ppb) |
|---------------------|--------------------|---------------------------------|------------------|------------|-----------------------------------|---|-------------------------|------------------------------------|
| VOC | 1 | Toluene | Т | 108-88-3 | C ₇ H ₈ | 92.1 | 330 | 10,000 |
| | 2 | p-Xylene | p-X | 106-41-3 | C ₈ H ₁₀ | 106.0 | 58 | 1000 |
| | 3 | Styrene | S | 100-42-5 | C_8H_8 | 104.0 | 35 | 400 |
| | 4 | Methyl ethyl ketone | MEK | 78-93-3 | C ₈ H ₈ O | 72.1 | 440 | 13,000 |
| | 5 | Isobutyl alcohol | i-BuAl | 78-83-1 | $C_4H_{10}O$ | 74.1 | 11.00 | 900 |
| | 6 | Methyl isobutyl ketone | MIBK | 108-10-1 | $C_{6}H_{12}O$ | 100.0 | 170 | 1000 |
| | 7 | Butyl acetate | BuAc | 123-86-4 | $C_6H_{12}O_2$ | 116.00 | 16 | 1000 |
| RSC | 8 | Hydrogen sulfide | H_2S | 7783-06-4 | H_2S | 34.1 | 0.41 | 20 |
| | 9 | Methyl mercaptan | CH₃SH | 74-93-1 | CH₃SH | 48.1 | 0.07 | 2 |
| | 10 | Dimethyl sulfide | DMS | 75-18-3 | (CH ₃) ₂ S | 62.1 | 3 | 10 |
| | 11 | Dimethyl disulfide | DMDS | 624-92-0 | $(CH_3)_2S_2$ | 94.2 | 2.2 | 9 |
| Aldehyde | 12 | Acetaldehyde | AA | 75-07-0 | C_2H_4O | 44 | 1.5 | 50 |
| | 13 | Propionaldehyde | PA | 123-38-6 | C_3H_6O | 58.1 | 1 | 50 |
| | 14 | Butyraldehyde | BA | 123-72-8 | C_4H_8O | 72.1 | 0.67 | 29 |
| | 15 | Isovaleraldehyde | IA | 590-86-3 | $C_5H_{10}O$ | 86.1 | 0.1 | 3 |
| | 16 | Valeraldehyde | VA | 110-62-3 | $C_5H_{10}O$ | 86.1 | 0.41 | 9 |
| Ν | 17 | Ammonia | NH ₃ | 7664-41-7 | NH ₃ | 17 | 1500 | 1000 |
| Relevant parameters | 18 | Formaldehyde | FA | 50-00-0 | CH ₂ O | 30 | 500 | - |
| | 19 | Benzene | В | 71-43-2 | C_6H_6 | 78.1 | 2700 | - |
| | 20 | Carbon disulfide | CS ₂ | 75-15-0 | CS_2 | 76.1 | 210 | - |
| | 21 | Sulfur dioxide | SO ₂ | 7446-09-5 | SO ₂ | 64.1 | 870 | - |
| | 22 | Carbon monoxide | CO | 630-08-0 | CO | 28 | - | - |
| | 23 | Total hydrocarbon | THC | - | - | - | - | - |
| | 24 | Particulate matter (10 μ g) | PM ₁₀ | | | - | - | - |

^a A total of five (trimethylamine and 4 volatile fatty acids (propionic acid, butyric acid, valeric acid, and isovaleric acid)) out of 22 offensive odorants were not considered; their concentrations were below detection limits from all samples.

^b Compounds of order 1–17 are offensive odorants designated by the malodor prevention law in Korea (KMOE, 2008) [13].

^c Permissible emission concentration of offensive odorants in non-industrial sectors according to the malodor prevention law in Korea (KMOE, 2008) [13].

products were combusted in an old-style Korean combustor (sample weight: ~550 g). Gases released from charcoal combustion were collected in 10 L Tedlar bags using a lung vacuum sampler (ACEN Co. Ltd., Korea). After releasing the first 5 min of gas emissions, gas samples were collected for 30 min into the Teflon bag placed in the lung sampler through a sampling line (made of Teflon) connected to the chimney of the combustor. These samples collected in the Tedlar bag were then used for the analysis of VOC and RSC.

For the collection of carbonyls, charcoal fumes were drawn into Lp-DNPH cartridges (Supelco Inc., USA) at a flow rate of 1 Lmin^{-1} for 5 min via a Sep-Pak ozone scrubber (Waters, USA). In the case of ammonia (NH₃), gaseous samples were absorbed directly into a boric acid solution made by 2.5 g boric acid mixed into 500 mL. To induce absorption of NH₃, three impinges were connected to each other; two of them were filled with boric acid (20 mL each), while the other was filled with resin (for dehydration of outgoing air). Next, a vacuum pump (MP- Σ 300, SIBATA, Japan) was used to draw gas samples through impinger at a flow rate of 2.5 L min⁻¹ to make a total of 30 L.

2.2. Analysis of odorant components

All collected samples were analyzed to determine concentration of up to 22 major offensive odorants designated by the malodor prevention law in Korea (KMOE, 2008) along with other reference pollutants (benzene, CS₂, SO₂, CO, THC, and PM₁₀) (Table 1). The concentration data of 5 of the 22 offensive odorants (trimethyl amine, propionic acid, butyric acid, valeric acid, and isovaleric acid) were not evaluated due to their limited detectabilty. Instead, a list of reference compounds was selected in this study and analyzed due to their relative abundance, irrespective of their odorant intensities. The basic analytical settings and the basic experimental conditions for their analysis are described in Table 2. The analysis of these odorants was made by following the procedures introduced in a number of previous investigations [5,9,10]. The basic information for these procedures is described briefly below.

For VOC analysis, the combination of gas chromatography (GC) with mass spectrometry (MS) coupled with a multifunction thermal desorber (TD) was used. The samples in the Tedlar bag were transported to the TD system for analysis based on the thermal desorption. Chromatographic separation was achieved by the Vocol column ($60 \text{ m} \times 0.32 \text{ mm}$ i.d. and 1.8-µm film thickness: Supelco) at a column flow rate $1.2 \text{ mL} \text{min}^{-1}$ (99.9% pure He as carrier gas). Detailed analytical conditions of this system are listed in Table 2. The detection limit (DL) values fell in the range of 1.27 (0.31 ppb (MIBK)) to 1.81 ng (0.38 ppb (BuAc)). If the precision of this method is evaluated in terms of relative standard error (RSE), it generally varied from 2.96% (MEK) to 4.96% (BuAc).

The analysis of RSC was made by GC with pulsed flame photometric detector (PFPD), which is interfaced with a multi-function TD and an air server (AS) unit. Detailed operating conditions of this system are described in Table 2. The analytical procedures of RSC can also be referred to in some of previous publications [11,12]. The DL values of RSC ranged from of 0.5 pg (or 0.12 ppb (DMDS)) to 0.7 pg (or 0.52 ppb (H₂S)) in a sampling volume of 120 mL. Their RSC values ranged from 1.35 (H₂S) to 4.25% (DMDS).

For carbonyl analysis, high performance liquid chromatography (HPLC) equipped with a UV detector and dsCHROM software (for peak integration) was used. The basic analytical conditions of the HPLC system are provided in Table 2. The RSE values were in the range of 0.51% (AA) to 2.16% (IA). The DL of the carbonyls ranged from 0.04 (VA) to 0.52 ppb (FA). Furthermore, NH₃-absorbed boric acid was analyzed by the UV/VIS detector system based on the colorimetric method (Table 2). Quantification of NH₃ was carried out with a DL value of 133 ppb and RSE of 1.95%. Due to problems with sample handling, quantification of some VOCs (e.g., MEK, i-BuAl, MIBK, and BuAc) was confined to three Korean samples and one US sample (K1–K3 and U1) (Table 3). Apart from the target odorants, some reference components (PM₁₀, CO, and THC) were also analyzed in light of their abundance in fume samples.

Table 2

| Exi | perimental | conditions | of the inst | rumental s | vstems use | d for o | dorant o | detection | in this s | tudv. |
|-----|------------|------------|-------------|------------|------------|---------|----------|-----------|-----------|-------|
| | | | | | <i></i> | | | | | |

| (a) Oven condition (b) Detector (PFPD) (c) Column (Vacol, PA, USA) (c) Column (Pariser (| [1] GC/MS system for VOC anal 1. GC/MS (SHIMADZU GCMS-Q | ysis P2010, Japan) | | | | |
|---|--|---|----------------------|--------------------------------------|------------------|----------------------|
| Initial temp: 35 "C Ionization mode: EI (70 eV) Hold time: 4 min Ionisaurce temps: 2000 C Ramping rate: 4 'Cmin' ⁻¹ TC scan range: 35-250 m/z Final temp: 200 'C 'The scan range: 35-250 m/z Gartier gas: He 99.90% '''''''''''''''''''''''''''''''''''' | (a) Oven condition | | | (b) Detector (MS) | | |
| Hold time: 4 min Ion source temps: 200 °C Ramping rate: 4 'Cmin-1 TICs can range: 35–250 m/z Final temp: 200 'C Threshold: 100 m/z Garrier gas: He 99.90% - | Initial temp.: | 35 | °C | Ionization mode: | EI (70 eV) | |
| Ramping rate:4'C min ⁻¹ TC scan range:35–250m/zFinal temp:200'CThreshold:100Cald time:10minCarl trags:He99.90%Carl trags:He99.90%Carl trags:Kernel Alexan range:Column (Vocol, PA, USA)0.32mmColumn (Vocol, PA, USA)60mmmmLength:Column (Vocol, PA, USA)60mm2. Thermal desorber (UNITY, Markers International Ltd., UK)Column (Vocol, PA, USA)0.32mm2. Thermal desorber (UNITY, Markers International Ltd., UK)Trap high:300CCold trags:5.0mL min ⁻¹ Flow path temperature:120CSplit ratio:20mL min ⁻¹ Flow path temperature:220C11.CC/PPPO (Do 6500, Donar Intrument, Korea) system(b) Detector (PPP): Model 5380, OL Analytical, USA)min11.CC/PPPO (Do 5600, Donar June)29.0'Cmin(a) Oven condition80'CMinAirt () Alfor:10min(a) Oven condition200'C'Cmin(c) Column (PP-1, SGE, Australia)minInitial temp:200'C'CminMin(c) Column (PP-1, SGE, Australia)minInitial temp:200'C'C'TminDiameter:0.32minInitial temp:200'C'C'T'TminDiameter:0.32minInitial temp:20'C< | Hold time: | 4 | min | Ion source temp.: | 200 | °C |
| Final remp: 200 "C The Indication of C The Indication of C The Indication of C Hold time: 10 min Indication of C Indication of C Indication of C Carrier gas: He 99.90% Indication of C Indicatio C | Ramping rate: | 4 | °C min ⁻¹ | TIC scan range: | 35-250 | m/z |
| Hold time: Carrier gas:10min 99.0%Carrier gas:He99.0%() Column (Vocol, PA, USA) Column (Vocol, PA, USA)0.32m m Length: 60mCold tap:Carbopack B' Tenaxran bickness:1.8Cold tap:Carbopack B' TenaxTrap high: | Final temp: | 200 | °C | Threshold: | 100 | |
| Carrier gas:He99.90%Carrier gas:He99.90%Column (Vocol, PA, USA)0.32mmColumn (Vocol, PA, USA)0.32mmEngit:60mFilm thickness:1.8mm2. Thermal desorber (UNITY, Markers International Ltd, UK)Trap low:5Split flow:5.0mL min ⁻¹ Film thickness:30.0Split flow:5.0mL min ⁻¹ Film thickness:12.0Hold time:5.0mL min ⁻¹ Film thickness:5(1) Core condition5.0mL min ⁻¹ Film thickness:5(2) Core condition0CDetector (PFPD: Model 5380, OL) Analytical, USA)mL min ⁻¹ Initial temp:200°C min ⁻¹ Air(1)/air(2): flow:10.0mL min ⁻¹ Final temp:200°C min ⁻¹ Air(1)/air(2): flow:11.5mL min ⁻¹ Initial temp:20minCMeteror remp:32.0mTotal time:2.0minCMeteror remp:3.2m11.5minCMeteror remp:3.2m2. Internal desorber (UNITY, Marker stational Ltd, UK)Trap logh:25.0mCSplit ratio:15.0mL min ⁻¹ Trap logh:25.0CCSplit ratio:15.0mL min ⁻¹ Pore staze:5.0mCFilm Hickness:5mLMetheressice5.0mCColuma (Classice) Classice) Classice) Classice< | Hold time: | 10 | min | | | |
| the function of the section of the | Carrier gas: | He | 99 90% | | | |
| $ \begin{array}{c c c c c c } Column (Vocol, PA, USA) & 0.32 & mm & 0.32 & mm$ | 8 | | | (c) Column (Vocol PA USA) | | |
| $ \begin{array}{ c c c c c c } \mbox{Length:} & 60 & m \\ \mbox{Length:} & 60 & m \\ \mbox{Line matrix} & 1.8 & \mu m \\ \mbox{Line matrix} & 20 & Trap high: & 30 & C \\ \mbox{Split ratio:} & 20 & m \\ \mbox{Line matrix} & 5.0 & C \\ \mbox{Line matrix} & 5.0 & C \\ \mbox{Line matrix} & 5.0 & C \\ \mbox{Line matrix} & 0.0 & C & Detector (PFPD: Model 5380, OL Analytical, USA) \\ \mbox{Line matrix} & 10 & m \\ \mbox{Line matrix} & 10 & m \\ \mbox{Line matrix} & 20 & C \\ \mbox{Ramping rate:} & 20 & C & min \\ \mbox{Line matrix} & 20 & C & min \\ \mbox{Line matrix} & 10 & m \\ \$ | | | | Column (Vocol PA LISA) | 0.32 | mm |
| Image: the second of the se | | | | Length: | 60 | m |
| 2. Thermal desorber (UNITY, Markers International Ltd., UK) Trap low: 5.0 Trap high: 300 °C Split ratio: 20 Split ratio: 5.0 mLmin ⁻¹ Flow path temperature: 120 °C Hold time: 5.0 ml.min ⁻¹ Flow path temperature: 120 °C Split ratio: 5.0 ml.min ⁻¹ Flow path temperature: 120 °C Split ratio: 5.0 ml.min ⁻¹ Flow path temperature: 120 °C C Split ratio: 5.0 ml.min ⁻¹ Flow path temperature: 250 °C ml.min ⁻¹ flow path temperature: 250 °C Ramping rate: 20 °C min ⁻¹ Air(1)/air(2): flow: 10 ml.min ⁻¹ Flow path temperature: 11.5 ml.min ⁻¹ Initial temp: 200 °C Hg flow: 11.5 ml.min ⁻¹ Flint Hickness: 5.0 min (c) Column (BP-1, SGE, Australia) Frial temp: 200 °C Hg flow: 11.5 ml.min ⁻¹ Flint Hickness: 5.0 min (c) Column (BP-1, SGE, Australia) Total time: 20 min Flint the set in ternational Ltd., UK) Column desorber (UNITY, Markers International Ltd., UK) Column desorber (UNITY, Markers International Ltd., UK) Column desorber (UNITY, Markers International Ltd., UK) Trap high: 250 °C Split ratio: 10:01 Trap high: 250 °C Split ratio: 150 ml.min ⁻¹ Flow path temperature: 80 °C Split ratio: 150 ml.min ⁻¹ Flow path temperature: 80 °C Split ratio: 150 ml.min ⁻¹ Flow path temperature: 55 mm flim the set in temperature: 50 ml.min ⁻¹ Flow path temperature: 250 °C Ads mm flipter UV detector (Model Sol0, Lab Alliance, USA) UV system for carbonyl compounds analysis 13(Hg C) Set in the set in the set in ternational Ltd., UK (C) Column (C18, Hichrom, UK) COLUMN (C) Set in the set in ternational Ltd., UK (C) Trap high: 250 °C Mol A Monomeric Column (C18, Hichrom, UK) CO | | | | Film thickness: | 18 | um |
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| $ \begin{array}{c c c c c } \mbox{Jpm ratio} & 20 & \mbox{Imm}^{-1} & \mbox{Imp} ratio} & \mbox{Jpm ratio} & Jpm $ | Split ratio: | | | Trap high: | 300 | °C |
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| [2] GC/TD system for RSC and Jistrament, Korea) system (A) Oven condition (B) Detector (PFPD: Model 5380, U. L-alytical, USA) [a) Oven condition (b) Detector (PFPD: Model 5380, U. L-alytical, USA) Vector [a) Main condition (C) Detector (PFPD: Model 5380, U. L-alytical, USA) (C) [a) Main condition (C) Condition (C) (C) [a] Main condition (C) Condition (C) (C) (C) [a] Main condition (C) Column (BP-1, SCE, Australia) (C) (C) (C) [a] Main condition (C) Column (BP-1, SCE, Australia) (C) (C) (C) (C) [a] Main condition (C) Column (BP-1, SCE, Australia) (C) (C) (C) (C) [a] Main condition (C) Column (C) (C) (C) (C) (C) [a] Main condition (D) (C) (C) | Hold time: | 5.0 | 111111 | | | |
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| Ramping rate:20°C min^{-1}Air(1)/air(2): flow:10mLmin^{-}Final temp:200°CH2 flow:11.5mLmin^{-}Initial hold:4.5min | Initial temp.: | 80 | °C | Detector temp.: | 250 | °C |
| Final temp:200°CH2 flow:11.5mLminInitial hold:4.5min11.5mLminFinal hold:9.5min(°C Column (BP-1, SGE, Australia))Total time:20minFilm thickness:5µmTotal time:20minFilm thickness:50µm2. Thermal desorber (UNITY, Markers International Ltd., UK)60mm2. Thermal desorber (UNITY, Markers International Ltd., UK)0.32mm2. Thermal desorber (UNITY, Markers International Ltd., UK)-15°CCold trap:Carbopack B* Silica gel = 1.5:2.5Trap low:-15°CSplit flow:15mLmin ⁻¹ Flow path temperature:80.0°CSplit flow:5mL min ⁻¹ Flow path temperature:80.0°CHold time:5mL min ⁻¹ Flow path temperature:5.0µmInjectorColumn (C18, Hichrom, UK)Volume:250 × 46mmVolume:20µLColumn dimensions:250 × 46mmPumpParticle size:300AAMobile phase:1.5mL min ⁻¹ Pore size:300AMobile phase:360mmImminImminImmin[4] UV/VIS Spectrometer (Genestrate, Thermo Electron Core, USA) system for ammonia-malysisImminImminImpringer system2.5mL min ⁻¹ Detector:UV/VIS | Ramping rate: | 20 | °C min ⁻¹ | Air(1)/air(2): flow: | 10 | mL min ⁻¹ |
| Initial hold:4.5minFinal hold:9.5min(c) Column (BP-1, SCE, Australia)Total time:20minFilm thickness:5 μ mLength:60mLength:0.32mm2. Thermal desorber (UNITY, Markers International Ltd., UK)Image: Column (BP-1, SCE, Australia)mm2. Thermal desorber (UNITY, Markers International Ltd., UK)0.32mm2. Thermal desorber (UNITY, Markers International Ltd., UK)Column (Clampic Column (Cla | Final temp: | 200 | °C | H ₂ flow: | 11.5 | mL min ⁻¹ |
| Final hold:9.5min(c) Column (BP-1, SGE, Australia)Total time:20minFilm thickness:5 μ mTotal time:20minFilm thickness:60min2. Thermal desorber (UNITY, Markers International Ltd., UK)Image: Column (BP-1, SGE, Australia)mm0.32mm2. Thermal desorber (UNITY, Markers International Ltd., UK)Trap low:0.32mmColumn (BP-1, SGE, Australia)mm2. Thermal desorber (UNITY, Markers International Ltd., UK)Trap low:15Column (BP-1, SGE, Australia)Column (BP-1, SGE, Aus | Initial hold: | 4.5 | min | - | | |
| Total time:20minFilm thickness:5 μ mTotal time:20minFilm thickness:60mLength:0.32mm2. Thermal desorber (UNITY, Markers International Ltd., UK)Diameter:0.32mmCold trap:Carbopack B* Silica gel=1.5:2.5Trap low:-15°CSplit ratio:10:01Trap high:250°CSplit flow:15mL min ⁻¹ Flow path temperature:80°CHold time:5min°C[3] HPLC (Series 1500, Lab Alliance, USA)/UV system for carbonyl compounds analysisColumn (C18, Hichrom, UK)°CVolume:20 μ LColumn dimensions:250 × 46mmPump250 × 06mmParticle size:5 μ mFlow rate:1.5mL min ⁻¹ Pore size:300AMobile phase:Acetronitrile; water70:30Temp:20°CAnalysis time:15minTemp:MonomericUVUV detector (Model 500, Lab Alliance, USA)mmWavelength:360mmmmImpringer systemImpringer systemUV/VISPump flow rate:2.5LML min ⁻¹ Detector:UV/VIS | Final hold: | 9.5 | min | (c) Column (BP-1, SGE, Australia) | | |
| $\begin{tabular}{ c c c c c } \hline lent time in the initial constraint of the initial constraint o$ | Total time: | 20 | min | Film thickness: | 5 | um |
| $\begin{tabular}{ c c c c } \hline lend to the term of ter$ | rotar timer | 20 | | Length: | 60 | m |
| 2. Thermal desorber (UNITY, Markers International Ltd., UK) Cold trap: Carbopack B* Silica gel = 1.5:2.5 Trap low: -15 °C Split ratio: 10:01 Trap high: 250 °C Split flow: 15 mL min ⁻¹ Flow path temperature: 80 °C Hold time: 5 Column (C18, Hichrom, UK) [3] HPLC (Series 1500, Lab Alliance, USA)/UV system for carbonyl compounds analysis Injector Column (C18, Hichrom, UK) Volume: 20 µL Column dimensions: 250 × 46 mm Pump Particle size: 5 µm Flow rate: 1.5 mL min ⁻¹ Pore size: 300 A Mobile phase: Acetronitrile:water 70:30 Temp: 20 °C Analysis time: 15 min Temp: Monomeric UV detector (Model 500, Lab Alliance, USA) Wavelength: 360 mm [4] UV/VIS Spectrometer (GenesysTM 10 series, Thermo Electron Corp., USA) system for anmonia analysis Impringer system Pump fow rate: 2.5 mL min ⁻¹ Detector: UV/VIS | | | | Diameter: | 0.32 | mm |
| Carbonack B* Silica gel= 1.5:2.5Trap low: Trap high:-15CSplit ratio:10:01Trap high:250°CSplit flow:15mLmin ⁻¹ Flow path temperature:80°CHold time:5minColumn (C18, Hichrom, UK)'[3] HPLC (Series 1500, Lab Alliance, USA)/UV system for carbonyl compounds analysisColumn (C18, Hichrom, UK)Volume:250 × 46mm'[3] HPLC (Series 1500, Lab Alliance, USA)/UV system for carbonyl compounds analysisColumn (C18, Hichrom, UK)Volume:250 × 46mm'Volume:20 μ LColumn (C18, Hichrom, UK)Volume:250 × 46mmPump Flow rate:1.5mLmin ⁻¹ Pore size:5 μ mFlow rate:1.5mLmin ⁻¹ Pore size:300AMobile phase:Acetronitrile:water70:30Temp:20°CAnalysis time:15minTemp:MonomericUV detector (Model 500, Lab Alliance, USA)mmImpringer systemImpringer systemImpringer systemImpringer systemPump flow rate:2.5mLmin ⁻¹ Detector:UV/VIS | 2. Thermal desorber (UNITY_M | arkers International Ltd LIK) | | Diameteri | 0.02 | |
| Split ratio: 10:01 Trap high: 250 °C Split ratio: 10:01 Trap high: 250 °C Hold time: 5 mL min ⁻¹ Flow path temperature: 80 °C Hold time: 5 ml min ⁻¹ Flow path temperature: 80 °C Hold time: 5 ml min ⁻¹ Flow path temperature: 80 °C (3] HPLC (Series 1500, Lab Alliance, USA)/UV system for carbonyl compounds analysis Injector Column (C18, Hichrom, UK) Volume: 20 μ L Column dimensions: 250 × 46 mm Pump Particle size: 5 μ m Flow rate: 1.5 mL min ⁻¹ Pore size: 300 A Mobile phase: Acetronitrile:water 70:30 Temp: 20 °C Analysis time: 15 min Temp: Monomeric UV detector (Model 500, Lab Alliance, USA) Wavelength: 360 mm [4] UV/VIS Spectrometer (GenesysTM 10 series, Thermo Electron Corp., USA) system for ammonia analysis Impringer system Pump flow rate: 2.5 mL min ⁻¹ Detector: UV/VIS | Cold trap: | Carbonack B^+ Silica gel = 1.5.2.5 | | Tran low: | -15 | °C |
| Split ratio.10.0111.01< | Split ratio: | 10·01 | | Trap high: | 250 | °C |
| Spin low.15InfinitTow part chiperature.60CHold time:5min10CCC[3] HPLC (Series 1500, Lab Alliance, USA)/UV system for carbonyl compounds analysisInjectorColumn (C18, Hichrom, UK)Volume:20 μ LColumn dimensions:250 × 46mmPumpParticle size:5 μ mFlow rate:1.5mL min ⁻¹ Pore size:300AMobile phase:Acetronitrile:water70:30Temp:20°CAnalysis time:15minTemp:MonomericUVUV detector (Model 500, Lab Alliance, USA)mmImpringer system[4] UV/VIS Spectrometer (GenesysTM 10 series, Thermo Electron Corp., USA) system for analysisImpringer systemImpringer systemUV/VISPump flow rate:2.5mL min ⁻¹ Detector:UV/VISUV/VIS | Split flow: | 15 | mI min ⁻¹ | Flow path temperature: | 80 | °C |
| Find time.5Imm[3] HPLC (Series 1500, Lab Alliance, USA)/UV system for carbonyl compounds analysisInjectorColumn (C18, Hichrom, UK)Volume:20μLColumn dimensions:250 × 46PumpParticle size:Flow rate:1.5Mobile phase:Acetronitrile:water70:30Temp:Analysis time:1515minUV detector (Model 500, Lab Alliance, USA)Wavelength:360a60mm[4] UV/VIS Spectrometer (GenesysTM 10 series, Thermo Electron Corp., USA) system for ammonia analysisImpringer systemPump flow rate:2.5Num flowDetector:UV (VIS | Jold times | 15 | min | now path temperature. | 80 | C |
| $ \begin{array}{c c c c c } 1 \mbox{Pull} L(Series 1500, Lab Allisure, USA)/UV system for carbonyl compounds analysis \\ lnjector & Column (C18, Hichrom, UK) \\ \hline Volume: & 20 & \mu L & Column dimensions: & 250 \times 46 & mm \\ Pump & Particle size: & 5 & \mum \\ Flow rate: & 1.5 & mL min^{-1} & Pore size: & 300 & A \\ Mobile phase: & Acetronitrile:water & 70:30 & Temp: & 20 & °C \\ Analysis time: & 15 & min & Temp: & Monomeric \\ UV detector (Model 500, Lab Allisure, USA) \\ Wavelength: & 360 & mm \\ \hline I 4] UV/VIS Spectrometer (GenessTM 10 series, Thermo Electron Corp., USA) system for ammonistrational substance \\ Impringer system \\ Pump flow rate: & 2.5 & mL min^{-1} & Detector: & UV/VIS \\ \end{array} $ | Hold tille. | 5 | 111111 | | | |
| $\begin{tabular}{ c c c c } \label{eq:linear} Index I$ | [3] HPLC (Series 1500, Lab Allia | nce, USA)/UV system for carbonyl compou | nds analysis | | | |
| $ \begin{array}{cccccccccccccccccccccccccccccccccccc$ | Injector | | | Column (C18, Hichrom, UK) | | |
| $\begin{array}{c c c c c } Pump & & Particle size: & 5 & \mum \\ Flow rate: & 1.5 & mL min^{-1} & Pore size: & 300 & A \\ Mobile phase: & Acetronitrile:water & 70:30 & Temp: & 20 & °C \\ Analysis time: & 15 & nin & Temp: & Monomeric & °C \\ Analysis time: & 15 & nin & Temp: & Monomeric & °C \\ UV detector (Model 500, Lab Alliance, USA) \\ UV detector (Model 500, Lab Alliance, USA) \\ Wavelength: & 360 & mm & & & & & & \\ \hline [4] UV/VIS Spectrometer (Genesser M 10 series, Thermo Electron Corp., USA) system for ammonis USA \\ Impringer system & & & & & & & \\ \hline Pump flow rate: & 2.5 & mL min^{-1} & Detector: & UV/VIS \\ \end{array}$ | Volume: | 20 | μL | Column dimensions: | 250 	imes 46 | mm |
| Flow rate:1.5mL min^{-1}Pore size:300ÅMobile phase:Acetronitrile:water70:30Temp:20°CAnalysis time:15minTemp:MonomericUV detector (Model 500, Lab Alliance, USA) Wavelength:360mm-[4] UV/VIS Spectrometer (GenesysTM 10 series, Thermo Electron Corp., USA) system for ammonia analysis Impringer system Pump flow rate:2.5mL min^{-1}Detector:UV/VIS | Pump | | | Particle size: | 5 | μm |
| Mobile phase:Acetronitrile:water70:30Temp:20°CAnalysis time:15minTemp:MonomericUV detector (Model 500, Lab Alliance, USA)Wavelength:360mm[4] UV/VIS Spectrometer (GenesysTM 10 series, Thermo Electron Corp., USA) system for annalysisImpringer systemPump flow rate:2.5mL min^{-1}Detector:UV/VIS | Flow rate: | 1.5 | mL min ⁻¹ | Pore size: | 300 | A |
| Analysis time: 15 min Temp: Monomeric UV detector (Model 500, Lab Alliance, USA) Wavelength: 360 mm Imprine Imprine [4] UV/VIS Spectrometer (GenesysTM 10 series, Thermo Electron Corp., USA) system for ammonia analysis Impringer system UV/VIS UV/VIS Pump flow rate: 2.5 mL min ⁻¹ Detector: UV/VIS | Mobile phase: | Acetronitrile:water | 70:30 | Temp: | 20 | °C |
| UV detector (Model 500, Lab Alliance, USA) Wavelength: 360 mm [4] UV/VIS Spectrometer (GenesysTM 10 series, Thermo Electron Corp., USA) system for ammonia analysis Impringer system Pump flow rate: 2.5 mL min ⁻¹ Detector: UV/VIS | Analysis time: | 15 | min | Temp: | Monomeric | |
| Wavelength: 360 mm [4] UV/VIS Spectrometer (GenesysTM 10 series, Thermo Electron Corp., USA) system for ammonia analysis Impringer system Pump flow rate: 2.5 mL min ⁻¹ Detector: UV/VIS | UV detector (Model 500, Lab Al | lliance, USA) | | r r | | |
| [4] UV/VIS Spectrometer (GenesysTM 10 series, Thermo Electron Corp., USA) system for ammonia analysis Impringer system Pump flow rate: 2.5 mL min ⁻¹ Detector: UV/VIS | Wavelength: | 360 | mm | | | |
| [4] UV/VIS Spectrometer (GenesysTM 10 series, Thermo Electron Corp., USA) system for ammonia analysis Impringer system Pump flow rate: 2.5 mL min ⁻¹ Detector: UV/VIS | | | | | | |
| Impringer system Pump flow rate: 2.5 mL min ⁻¹ Detector: UV/VIS | [4] UV/VIS Spectrometer (Gene | esysTM 10 series, Thermo Electron Corp., US | SA) system for ammo | nia analysis | | |
| Pump flow rate: 2.5 mL min ⁻¹ Detector: UV/VIS | Impringer system | | | | | |
| | Pump flow rate: | 2.5 | mL min ⁻¹ | Detector: | UV/VIS | |
| Volume absorbed:30LWavelength:635nm | Volume absorbed: | 30 | L | Wavelength: | 635 | nm |
| Absorption time: 12 min | Absorption time: | 12 | min | | | |
| Temperature: 22 °C | Temperature: | 22 | °C | | | |
| Boric acid volume: 50 mL | Boric acid volume: | 50 | mL | | | |

3. Results and discussion

3.1. General features of odorant pollutant emission during combustion

The emission concentrations of odorant pollutants with diverse chemical properties were measured from 15 different charcoal products made in five different countries (Table 3). A brief inspection of the data indicates that the magnitude of odorant emissions differs significantly between compounds and between individual charcoals. According to the statistical summary provided in Table 4, the mean values of AA, PA, IA, and ammonia exceeded the guideline of malodor prevention law in Korea [13]. Their mean concentrations were almost 20 (1009 ppb), 7 (354 ppb), 2 (6.35 ppb), and 7 times (7800 ppb) higher than their respective guideline values. In contrast, emissions of certain species (RSCs, VA, etc.) were commonly limited in their delectability in most samples (Table 3).

Although VOCs are the key components of charcoal combustion, their concentrations are greatly distinguished between samples.

Among VOCs, p-X was measured consistently above DL in all charcoal products except for the one made in the US. Although benzene is not an odorant, its concentration levels from glowing charcoals have been measured frequently [14]. Due to the carcinogenic effect and the susceptibility of inhalation [15], benzene is designated as one of the top-priority compounds for the development of indoor air quality guidelines [16]. In this study, the concentration (ppb) of benzene averaged at 843, which was far higher than toluene (4.07). This contrasts with a comparable trend between toluene (116) and benzene (98.7) in previous study [4]. In the case of RSCs, their emission concentrations were generally low compared to VOCs and aldehydes. Although odorant RSCs were seen abundantly from a Korean sample (K2), they were not measured from any other sample. In contrast, the emission concentrations of other sulfur species (CS₂ and SO₂) from our samples were relatively high with averages of 3.85 and 14.9 ppb, respectively. In this research, a total of five odorant carbonyl (AA, PA, BA, IA, and VA) were measured along with FA as a reference. Among aldehydes, the concentrations (ppb) of AA (1009) and FA (354) were generally the highest, while the

 Table 3

 Emission concentrations (ppb) of target odorants and reference compounds measured from charcoal combustion experiments in this study.

| Group | Order | Short name ^a | Sample co | de ^b | | | | | | | | | | | | | |
|---------------------|-------|-------------------------------|-----------|-----------------|---------|--------------|-----------------|--------------|-----------|--------------|-----------|-----------|------------|--------------|-----------|-----------|------|
| | | | K1 | K2 | K3 | K4 | K5 | K6 | C1 | C2 | C3 | C4 | I1 | I2 | I3 | M1 | U1 |
| VOC | 1 | Т | 21.8 | 14.1 | 15.5 | 0.94 | <u>0.0003</u> ¢ | 0.0003 | 0.0003 | 0.0003 | 0.0003 | 0.0003 | 0.0003 | 0.0003 | 0.0003 | 0.65 | 8.10 |
| | 2 | p-X | 0.44 | 492 | 3.10 | 1.63 | 27.00 | 4.39 | 1.09 | 1.07 | 1.17 | 1.69 | 1.58 | 1.21 | 0.39 | 0.94 | 0.08 |
| | 3 | S | 6.73 | 7629 | 57.7 | 0.0005 | 10.0 | 0.0005 | 0.0005 | 0.0005 | 0.82 | 0.0005 | 0.0005 | 0.0005 | 0.0005 | 0.0005 | 0.06 |
| | 4 | MEK | 5.74 | 5915 | 108 | - | - | - | - | - | - | - | - | - | - | - | 0.17 |
| | 5 | i-BuAl | 0.26 | 2001 | 0.26 | - | - | - | - | - | - | - | - | - | - | - | 0.17 |
| | 6 | MIBK | 0.17 | 0.17 | 2.36 | - | - | - | - | - | - | - | - | - | - | - | 0.08 |
| | 7 | BuAc | 0.29 | 0.29 | 0.29 | - | - | - | - | - | - | - | - | - | - | - | 0.08 |
| RSC | 8 | H_2S | 0.03 | 3.38 | 0.03 | <u>0.036</u> | <u>0.036</u> | <u>0.036</u> | 0.036 | <u>0.036</u> | 0.036 | 0.036 | 0.036 | <u>0.036</u> | 0.036 | 0.036 | 0.25 |
| | 9 | CH₃SH | 0.03 | 14.4 | 0.03 | 0.024 | 0.024 | 0.024 | 0.024 | 0.024 | 0.024 | 0.024 | 0.024 | 0.024 | 0.024 | 0.024 | 0.23 |
| | 10 | DMS | 0.02 | 97.5 | 0.24 | 0.024 | 0.024 | 0.024 | 0.024 | 0.024 | 0.024 | 0.024 | 0.024 | 0.024 | 0.024 | 0.024 | 0.17 |
| | 11 | DMDS | 0.09 | 42.4 | 0.14 | 0.017 | 0.017 | 0.017 | 0.017 | 0.017 | 0.017 | 0.017 | 0.017 | 0.017 | 0.017 | 0.017 | 0.11 |
| Aldehyde | 12 | AA | 83.0 | 5764 | 1415 | 213 | 5734 | 189 | 883 | 47.1 | 68.4 | 110 | 198 | 241 | 107 | 81.4 | 0.44 |
| | 13 | PA | 5.07 | 347 | 80.91 | 0.05 | 388 | 0.05 | 0.05 | 0.05 | 0.05 | 0.05 | 0.05 | 0.05 | 14.1 | 0.05 | 0.44 |
| | 14 | BA | 0.46 | 89.1 | 71.6 | 0.05 | 132 | 12.3 | 0.05 | 0.05 | 0.05 | 0.05 | 0.05 | 0.05 | 0.05 | 0.05 | 0.45 |
| | 15 | IA | 0.47 | 47.6 | 21.5 | 0.04 | 24.7 | 0.04 | 0.04 | 0.04 | 0.04 | 0.04 | 0.04 | 0.04 | 0.04 | 0.04 | 0.56 |
| | 16 | VA | 0.48 | 21.4 | 0.48 | 0.04 | 0.04 | 0.04 | 0.04 | 0.04 | 0.04 | 0.04 | 0.04 | 0.04 | 0.04 | 0.04 | 0.49 |
| N | 17 | NH_3 | 22,319 | 83,325 | 8928 | <u>21</u> | <u>55</u> | 24 | <u>18</u> | <u>10</u> | 24 | 26 | <u>16</u> | <u>3</u> | <u>5</u> | 9 | 1614 |
| Relevant parameters | 18 | FA | 99.4 | 13.6 | 831 | 397 | 1137 | 436 | 138 | 174 | 198 | 299 | 194 | 902 | 153 | 340 | 0.52 |
| | 19 | В | 0.03 | 6,535 | 221 | 132 | 2093 | 1081 | 314 | 98.8 | 325 | 254 | 100 | 1120 | 59.3 | 283 | 21.7 |
| | 20 | CS ₂ | 2.07 | 40.0 | 0.15 | 1.84 | 3.34 | 1.22 | 1.43 | 0.95 | 0.65 | 1.52 | 1.54 | 0.94 | 0.53 | 1.44 | 0.07 |
| | 21 | SO ₂ | 39.7 | 21.9 | 8.95 | 6.48 | 20.5 | 9.35 | 2.51 | 12.1 | 26.0 | 14.9 | 4.43 | 37.6 | 0.83 | 3.54 | - |
| | 22 | CO | 1,020,000 | 1,050,000 | 537,000 | 399,375 | 3,612,038 | 1,316,450 | 1,662,743 | 999,473 | 2,621,175 | 1,838,183 | 901,390 | 1,727,745 | 1,309,458 | 1,525,533 | 750 |
| | 23 | THC ^d | 7300 | 87,200 | 500 | 212,760 | 2,034,143 | 1,730,000 | 169,093 | 500 | 199,360 | 161,325 | <u>500</u> | 983,400 | 500 | 84,280 | 500 |
| | 24 | PM ₁₀ ^e | 800 | 8889 | 24,889 | 3833 | 2833 | 3667 | 2000 | 2667 | 2667 | 2667 | 2000 | 2500 | 3000 | 500 | 717 |

^a Refer to Table 1 for full name.

^b Sample code: K (Korea); C (China); I (Indonesia); M (Malaysia); and U (US).

^c Underlined numbers denote concentration values below detection limit. DL values of a given species are occasionally different due to measurement intervals or changes in sample loading condition or sample treatments. Superscript letters d and e denote concentration units of ppbC and μ g m⁻³, respectively.

Table 4

Comparison of mean concentration values of odorants and reference components measured from 15 BBQ charcoal products.

| Group | Order | Short name ^b | All | Korea | China | Indonesia | Malaysia | US |
|---------------|-----------------|-------------------------------|-----------|-----------|---------------------|------------|------------|------|
| A. Offensive | odorants (unit: | ppb) ^a | | | | | | |
| VOC | 1 | Т | 4.07 | 8.71 | 0.0003 ^c | 0.0003 | 0.65 | 8.10 |
| | 2 | p-X | 35.9 | 88.2 | 1.26 | 1.06 | 0.94 | 0.08 |
| | 3 | S | 514 | 1284 | 0.21 | 0.0005 | 0.0005 | 0.06 |
| | 4 | MEK | 1507 | 2009 | - | - | - | 0.17 |
| | 5 | i-BuAl | 500 | 667 | - | - | - | 0.17 |
| | 6 | MIBK | 0.64 | 0.84 | - | - | - | 0.08 |
| | 7 | BuAc | 0.12 | 0.29 | - | - | - | 0.08 |
| RSC | 8 | H_2S | 0.25 | 0.58 | 0.036 | 0.036 | 0.036 | 0.25 |
| | 9 | CH ₃ SH | 0.98 | 2.41 | 0.024 | 0.024 | 0.024 | 0.23 |
| | 10 | DMS | 6.53 | 16.3 | 0.024 | 0.024 | 0.024 | 0.17 |
| | 11 | DMDS | 2.85 | 7.11 | 0.017 | 0.017 | 0.017 | 0.11 |
| Aldehyde | 12 | AA | 1009 | 2233 | 277 | 182 | 81.40 | 0.44 |
| | 13 | PA | 55.7 | 137 | 0.05 | 4.72 | 0.05 | 0.44 |
| | 14 | BA | 20.4 | 50.9 | 0.05 | 0.05 | 0.05 | 0.45 |
| | 15 | IA | 6.30 | 15.7 | 0.04 | 0.04 | 0.04 | 0.56 |
| | 16 | VA | 1.49 | 3.65 | 0.04 | 0.04 | 0.04 | 0.49 |
| N | 17 | NH ₃ | 7753 | 19,104 | <u>19.7</u> | <u>8.1</u> | <u>9.0</u> | 1614 |
| B. Relevant p | arameters (unit | :: ppb) | | | | | | |
| | 18 | FA | 354 | 486 | 202 | 416 | 340 | 0.26 |
| | 19 | В | 843 | 1677 | 248 | 426 | 283 | 21.7 |
| | 20 | CS ₂ | 3.85 | 8.11 | 1.14 | 1.0 | 1.44 | 0.03 |
| | 21 | SO ₂ | 14.9 | 17.8 | 13.87 | 14 | 3.54 | - |
| | 22 | CO | 1,368,062 | 1,322,477 | 1,780,393 | 1,312,864 | 1,525,533 | 375 |
| | 23 | THC ^d | 378,007 | 678,609 | 132,507 | 327,967 | 84,280 | 250 |
| | 24 | PM ₁₀ ^e | 4242 | 7485 | 2500 | 2500 | 500 | 717 |

^a In case all data for a given country are below detection limit (BDL), it was used directly for the computation of mean value; however, if the BDL data are mixed with the normal data, then one half of BDL was used to derive the mean value for each country.

^{b,c} Refer to Tables 1 and 3, respectively.

^d Concentration unit = ppbC.

^e Concentration unit = $\mu g m^{-3}$.

detection of other carbonyls (PA, BA, IA, and VA) were generally confined in most samples. Moreover, fairly high concentrations of ammonia were recorded from many samples (such as K-2, -1, -3, and U1), which were around 83, 22, 10 and 1.5 times higher than the guidelines [13]. As ammonia undergoes gas-to-particle conversion with atmospheric acids, it may be present in salt from in the atmosphere [17,18].

The concentrations of the major reference components (CO, THC, and PM_{10}) were measured consistently in most samples with the mean of 1368 ppm, 378 ppmC, and 4242 μ g m⁻³, respectively (Table 4). As the occupational exposure of CO is generally regulated in the range of 20 to 50 ppm [19,20], its value measured in this study was about 27–68 times higher than those guideline values. Hence, if ventilation is not maintained properly, charcoal combustion may cause severe health effects. As such, the prevalence of high PM₁₀ levels from charcoal combustion is also a worrying component [21].

3.2. Comparison of odor strengths between different charcoal products in terms of odor intensity

As odors released either directly or indirectly from human activities cause adverse health effects, they are often classified as the contaminants requiring regulation [22]. In North America and Europe, odors have been ranked as the major targets of public complaints to regulatory agencies [23]. Due to the complexity of odorant mixing and/or the delicacy of its detection by the human nose, a quantitative description of odor exposure is limited [24]. For this reason, the use of the odor intensity (OI) concept is an efficient approach as it provides a parallel means to evaluate concentration data of the perceived odorants through numerical conversions [9].

For the conversion of odorant concentrations into the OI values, empirical equations introduced by Nagata [25] were used in our study. The OI scaling of 0–6 can be distinguished as follows: 0 (no odor), 1 (very weak), 2 (weak), 3 (distinct), 4 (strong), 5 (very strong), and 6 (intolerable) [26]. However, as the OI values

were converted to negative values in some cases, such values were excluded from comparative analysis for the sake of simplicity. The OI values of offensive odorants derived via such conversions are compiled in Table 5.

In summary, our data suggests that the magnitude of OI values differs greatly among different charcoal products. If the OI values are compared between different charcoal products, aldehydes were found to be the most dominant odorant followed by VOC. Although FA and AA were detected abundantly from most charcoal samples, other carbonyls (PA, BA, and IA) were only found abundantly from 3 Korean samples e.g., (K-2, 3, and 5). It should be noted that MIBK and BuAc were not significant as an odorant, with OI values below zero. In contrast, NH₃ exhibited the highest OI value (5.58) in K-2 charcoal, while its values for K-1, -3, and U1 were 4.63, 3.97, and 2.72, respectively.

As a simple means to assess the overall contribution to odor formation by all target compounds, the OI values of each individual compound were bound together to derive the total odor strength in terms of the "sum of odor intensity (SOI)". For the derivation of the SOI term, the following equations were employed in this study [27]:

 $\begin{aligned} \text{SOI} &= \log \left(\Sigma 10^{\text{OI}(ith)} \right) = \log \left(10^{\text{OI}(ith)1} + 10^{\text{OI}(ith)2} + 10^{\text{OI}(ith)3} \right. \\ &+ \cdots + 10^{\text{OI}(ith)n} \end{aligned}$

where $OI(ith) = \log 10^{OI(ith)}$.

As the mixing of odorants is a complicated process, one should bear in mind that the derivation of SOI should be considered an arbitrary approach to assess one of the various mixing effects. In terms of SOI, the strength of the odor emission peaked in K-2 (5.66) followed by K-5 (4.73), K-1 (4.61), K-3 (4.43), C-1 (3.80). In the rest of the charcoal products, SOI values ranged from 3.14 to 2.51 (Table 5). In terms of odor strength classification into a 0–6 numerical scale by the American Society for Testing and Materials [26], the result of different charcoal products was arbitrarily

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Comparison of odor intensity (OI) for each individual charcoal product investigated in this work.

Sample code: K (Korea); C (China); I (Indonesia); M (Malaysia) and U (US).

Negative OI values are not considered.

| Order | Compound | Function ^a | Sample | code ^b | | | | | | | | | | | | | |
|---------------------|---------------------|----------------------------|-------------|-------------------|------|------|------|------|------|------|------|------|------|------|------|------|------|
| | | | K1 | K2 | K3 | K4 | K5 | K6 | C | 2 | ១ | C4 | Ħ | 12 | 13 | M1 | IJ |
| - | F | Y= 1.380 log X+ 4.60 | 2.31 | 2.05 | 2.10 | 0.42 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | ı | 1 | 0.20 | 1.71 |
| 2 | p-X | $Y = 1.570 \log X + 2.44$ | Ŭ | 1.96 | I | I | I | I | I | I | I | I | I | ı | I | I | I |
| ę | s | $Y = 0.790 \log X + 2.53$ | 0.81 | 3.23 | 1.55 | I | 0.95 | I | I | I | 0.09 | I | I | I | I | I | I |
| 4 | MEK | $Y = 1.850 \log X + 0.15$ | I | 1.58 | I | I | I | I | I | I | I | I | I | I | I | I | I |
| IJ. | i-BuAl | $Y = 0.790 \log X + 2.53$ | I | 2.77 | I | I | I | I | I | I | I | I | I | I | I | I | I |
| 9 | MIBK | $Y = 1.650 \log X + 2.27$ | I | I | I | I | I | I | I | I | I | I | I | I | I | I | I |
| 7 | BuAc | $Y = 1.140 \log X + 2.34$ | I | I | I | I | I | I | I | I | I | I | I | I | I | I | I |
| 8 | H_2S | $Y = 0.950 \log X + 4.14$ | I | 1.79 | I | I | I | I | I | I | I | I | I | I | I | I | I |
| 6 | CH ₃ SH | $Y = 1.250 \log X + 5.99$ | I | 3.69 | I | I | I | I | I | I | I | I | I | ı | I | I | I |
| 10 | DMS | $Y = 0.784 \log X + 4.06$ | I | 3.27 | 1.22 | 0.44 | 0.44 | I | I | I | I | I | I | 0.44 | I | I | I |
| 11 | DMDS | $Y = 0.985 \log X + 4.51$ | | 3.16 | 0.72 | I | I | I | I | I | I | I | I | ı | I | I | I |
| 12 | AA | $Y = 1.010 \log X + 3.85$ | 2.76 | 4.62 | 4.00 | 3.17 | 4.62 | 3.12 | 3.80 | 2.51 | 2.67 | 2.88 | 3.14 | 3.23 | 2.87 | 2.75 | I |
| 13 | PA | $Y = 1.010 \log X + 3.86$ | 1.53 | 3.40 | 2.76 | I | 3.44 | I | I | I | I | I | I | I | 1.99 | I | I |
| 14 | BA | $Y = 1.060 \log X + 4.23$ | I | 3.12 | 3.02 | I | 3.30 | 2.21 | I | I | I | I | I | I | I | I | I |
| 15 | IA | $Y = 1.350 \log X + 6.01$ | I | 4.22 | 3.76 | I | 3.84 | I | I | I | I | 0.07 | 0.07 | I | I | I | I |
| 16 | NH ₃ | Y= 1.670log X + 2.38 | 4.63 | 5.58 | 3.97 | I | 0.28 | I | I | I | I | I | I | ı | I | I | 2.72 |
| 17 | FA | $Y = 1.530 \log X + 1.59$ | 0.06 | I | 1.47 | 0.98 | 1.68 | 1.04 | 0.27 | 0.43 | 0.51 | 0.79 | 0.50 | 1.52 | 0.34 | 0.87 | I |
| | SOI | | 4.61 | 5.66 | 4.43 | 3.17 | 4.73 | 3.17 | 3.80 | 2.51 | 2.67 | 2.88 | 3.14 | 3.24 | 2.92 | 2.76 | 2.76 |
| ^a Nagata | [25], odor intensit | y (Y) and odorant concentr | ation (ppm) | (X). | | | | | | | | | | | | | |

divided into 4 groups by their magnitude (1) Very strong (K-2), (2) Strong (K-1, K-3, and K-5), (3) Distinct (K-4, K-6, C-1, I-1, and I-2), and (4) Weak for all the remaining charcoal. According to a compilation of the SOI values between different countries, the odor intensity of the Korean samples tends to be significantly high compared to others (e.g., Malaysia and US).

3.3. Comparison of odorants between charcoal products of different countries

As our measurements were made to explore odorant emission characteristics covering 15 products from five different countries, the data can be evaluated to assess the possible role of such criteria as the origin of the charcoal production. As seen by the plot of the averaged concentrations from all the experimental results, charcoal consumption appears to be a significant odorant emission process (Fig. 1). If the mean values of the major odorants are compared between countries, the values of the Korean charcoal products record the highest of all countries (Fig. 2). In many Korean charcoals, the concentration values of all odorants (except T, p-X, MEK, and DMDS, Table 5) were high enough to exceed their respective guideline emission values [e.g., 13]. If we look into the common trend of pollutant emissions between different countries (such as Korea, Chinese, Indonesia, and Malaysia samples), the acetaldehyde (AA) value was about 45, 6, 4, and 2 times higher than the Korean guideline value. In the case of ammonia, it was found in a very high concentration range in Korean and US charcoal; its values were about 20 and 2 times higher than the guideline value (Table 4). It is worth noting that ammonia was the only prime odorant released from the US charcoal product tested in this study, while it was not for the products of the other countries.

If we compare the odor strengths of charcoal products between different countries in terms of SOI values, Korean products recorded the highest SOI value of 4.30 followed by Indonesian (3.10), Chinese (2.97), Malaysian (2.76), and the US product (2.76) (Fig. 2). The SOI values of the Korean charcoal products showed a little complicated pattern between different odorants, while those of other countries (Chinese, Indonesian, and Malaysian charcoal products) appear to be dominated by the AA compound. However, the pattern of US products is quite exceptional, as ammonia was the single dominant odorant contributing to the SOI level. This observation thus suggests the possibility that such product underwent a sufficient cleaning process in the handling of the raw materials compared to other countries.

3.4. Factors affecting odorant emissions from charcoal

In order to learn more about the factors affecting the odorant emissions from various charcoal products, Pearson's correlation analysis was done using the concentration data of all odorants (Table 6). Concentrations of some odorants (MEK, i-BuAl, MIBK, H₂S, CH₃SH, DMS, IA, and VA) were excluded in the correlation analysis due to the limited detectability. If the results are compared by the level of statistical significance (*p* value) of 0.05 and 0.01, a total of 6 and 28, respectively, out of 94 matching pairs were found to be statistically significant. In our study, benzene displayed a good correlation with most of the compounds, while toluene and p-X were only correlated significantly with ammonia. Among the aldehydes, FA, AA, PA, and BA showed good correlations with each other, suggesting the similar nature of their source processes among different carbonyls. In most cases, odorants of VOC (P, p-X, and S) were highly correlated with aldehydes (AA, PA, and BA).

In order to assess the source of different odorants (and pollutants) in charcoal fume, both the manufacturing process and raw materials used for its production need to be investigated. The natural raw materials for charcoal production are based on wood, both

| Table 6 |
|---|
| Results of correlation analysis between target odorants investigated and reference components in this study. ^a |

| | | Т | РХ | S | DMDS | AA | PA | BA | NH ₃ | FA | В | CS ₂ | SO ₂ | CO | THC | PM10 |
|-----------------|----------------|--------|---------|-------------|---------|---------|---------|---------|-----------------|-------|---------|-----------------|-----------------|--------|-------|------|
| (a) Resu | lts | | | | | | | | | | | | | | | |
| т | $r^{\rm b}$ | 1 | | | | | | | | | | | | | | |
| 1 | p ^c | - | | | | | | | | | | | | | | |
| | N ^d | 15 | | | | | | | | | | | | | | |
| ΡX | r | 0.376 | | | | | | | | | | | | | | |
| IX | р | 0.168 | 1 | | | | | | | | | | | | | |
| | Ν | 15 | 15 | | | | | | | | | | | | | |
| S | r | 0.387 | 0.999** | 1 | | | | | | | | | | | | |
| 5 | р | 0.154 | 0.000 | - | | | | | | | | | | | | |
| | Ν | 15 | 15 | 15 | | | | | | | | | | | | |
| DMDS | r | 0.386 | 0.999** | 0.999** | 1 | | | | | | | | | | | |
| DINIDU | р | 0.155 | 0.000 | 0.000 | | | | | | | | | | | | |
| | Ν | 15 | 15 | 15 | 15 | | | | | | | | | | | |
| AA | r | 0.220 | 0.708** | 0.672** | 0.671** | 1 | | | | | | | | | | |
| | р | 0.431 | 0.003 | 0.006 | 0.006 | - | | | | | | | | | | |
| | Ν | 15 | 15 | 15 | 15 | 15 | | | | | | | | | | |
| PA | r | 0.220 | 0.666** | 0.629^{*} | 0.627** | 0.992** | 1 | | | | | | | | | |
| | р | 0.432 | 0.007 | 0.012 | 0.012 | 0.000 | | | | | | | | | | |
| | Ν | 15 | 15 | 15 | 15 | 15 | 15 | | | | | | | | | |
| BA | r | 0.268 | 0.497 | 0.459 | 0.456 | 0.930** | 0.944** | 1 | | | | | | | | |
| DIT | р | 0.334 | 0.059 | 0.085 | 0.088 | 0.000 | 0.000 | | | | | | | | | |
| | Ν | 15 | 15 | 15 | 15 | 15 | 15 | 15 | | | | | | | | |
| NHa | r | 0.608* | 0.958** | 0.962** | 0.962** | 0.633* | 0.595* | 0.449 | 1 | | | | | | | |
| 11113 | р | 0.016 | 0.000 | 0.000 | 0.000 | 0.011 | 0.019 | 0.093 | | | | | | | | |
| | Ν | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | | | | | | | |
| FA | r | - | - | - | - | 0.327 | 0.358 | 0.541* | - | 1 | | | | | | |
| | р | 0.550 | 0.388 | 0.325 | 0.319 | 0.234 | 0.190 | 0.037 | 0.290 | • | | | | | | |
| | Ν | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | | | | | | |
| в | r | 0.263 | 0.954** | 0.940** | 0.940** | 0.829** | 0.794** | 0.643** | 0.880** | 0.004 | 1 | | | | | |
| 2 | р | 0.344 | 0.000 | 0.000 | 0.000 | 0.000 | 0.000 | 0.010 | 0.000 | 0.987 | | | | | | |
| | Ν | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | | | | | |
| CS ₂ | r | 0.374 | 0.998** | 0.997** | 0.997** | 0.707** | 0.664** | 0.487 | 0.960** | - | 0.952** | 1 | | | | |
| 652 | р | 0.170 | 0.000 | 0.000 | 0.000 | 0.003 | 0.007 | 0.066 | 0.000 | 0.375 | 0.000 | | | | | |
| | Ν | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | | | | |
| SO ₂ | r | 0.437 | 0.165 | 0.159 | 0.160 | 0.163 | 0.186 | 0.124 | 0.293 | 0.190 | 0.243 | 0.173 | 1 | | | |
| 2 | р | 0.119 | 0.573 | 0.587 | 0.585 | 0.579 | 0.525 | 0.672 | 0.309 | 0.515 | 0.403 | 0.554 | • | | | |
| | Ν | 14 | 14 | 14 | 14 | 14 | 14 | 14 | 14 | 14 | 14 | 14 | 14 | | | |
| CO | r | - | - | - | - | 0.408 | 0.430 | 0.397 | - | 0.483 | 0.153 | - | 0.280 | 1 | | |
| | р | 0.131 | 0.825 | 0.724 | 0.722 | 0.131 | 0.110 | 0.143 | 0.566 | 0.068 | 0.586 | 0.852 | 0.333 | • | | |
| | Ν | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 14 | 15 | | |
| THC | r | - | - | - | - | 0.399 | 0.424 | 0.471 | - | 0.689 | 0.209 | - | 0.200 | 0.603* | 1 | |
| | р | 0.244 | 0.772 | 0.664 | 0.662 | 0.140 | 0.115 | 0.076 | 0.519 | 0.005 | 0.456 | 0.779 | 0.493 | 0.017 | | |
| | Ν | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 14 | 15 | 15 | |
| PM10 | r | 0.456 | 0.214 | 0.220 | 0.215 | 0.264 | 0.248 | 0.480 | 0.266 | 0.348 | 0.185 | 0.184 | - | - | - | 1 |
| | р | 0.087 | 0.443 | 0.431 | 0.441 | 0.341 | 0.372 | 0.070 | 0.338 | 0.204 | 0.510 | 0.511 | 0.691 | 0.385 | 0.678 | • |
| | Ν | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 14 | 15 | 15 | 15 |

(b) Summary of correlation analysis

| Sample source | Frequency of n | natching pairs at 2 significance levels | Total number of possible matching pairs |
|---------------|----------------|---|---|
| | 0.01 | 0.05 | |
| | 28 | 6 | 94 |

^a A total of 9 odorants (MEK, i-BuAl, MIBK, BuAc, H₂S, CH₃SH, DMS, IA, and VA) are not considered for the correlation analysis as most of the values are below detection limit.

^b Pearson's correlation coefficient.

^c Probability (2 tails significance).

^d No. of data.

* Correlation is significant at the 0.05 level (2-tailed).

** Correlation is significant at the 0.01 level (2-tailed).



Fig. 1. Comparison of the mean concentration of major odorants and reference components measured from 15 different charcoal products (unit of THC = ppbC).

hardwood and softwood. In addition, sawdust, wood shavings, fruit stones, nuts, nutshells, corn cobs, bark, cotton seeds, and similar products can also be employed. Supply of wood came from two sources, directly from the forest or from industrial waste [28]. In light of this fact, it is logical to infer that recycled industrial wood. if used for charcoal, may play a certain role in the emission of hazardous odorants from charcoal fumes. Moreover, in our investigation, few charcoal products produced significant amounts of tarry residue during combustion. Note that liquid and tarry residues were not fully removed by the process of carbonization during production [29]. In addition, according to an investigation by the US EPA, when pyrolysis is used to manufacture charcoal from wood, a variety of different organic compounds are released into the air, depending on the specific pyrolysis or burn conditions [30]. The content of volatile components is almost zero, if carbonization is achieved at high temperatures (around 1000 °C). However, the negative effect of such processes is that the production can fall about 25% compared to low temperature carbonization [31]. In light of evidence collected from several previous researches, it is evident that the notable differences in pollutant emission levels were affected by the combined effects of their manufacturing ingredients and processes [4,5].

3.5. Assessment of potential health risks of odorants from charcoal combustion

The smoke from biomass fuels (wood, agricultural waste, and dung) is an important source of indoor air pollution because large amounts of smoke inhalation may induce a health risks of a similar order of magnitude as the risk from tobacco smoke [32]. In this study, major hazardous pollutants from charcoal combustion (odorants) were identified to include toluene, xylene, styrene, formaldehyde, acetaldehyde, and ammonia. According to [33], such compounds have the potential to pose cancer and non-cancer risks when present in dwellings and public buildings.

Most of the aromatics seen abundantly in this study (toluene, xylene, and styrene) are known to cause adverse health effects such as fatigue, sleepiness, headache, throat irritation, and short term memory loss [34]. During recent research on styrene, it was also found that its inhalation at both high and low concentration levels can damage DNA repair genes and pose long term cancer risks [35]. Although the level of odorant RSCs was insignificant, considerable amounts of other sulfur species like CS_2 and SO_2 were found in our study. In many previous studies, the inhalation of CS_2 is reported to destabilize the mechanism of carbohydrate metabolism in the human body [36–38]. In addition, SO_2 is known to exert certain effects on pulmonary function even at low concentrations [39].

A number of aldehydes, especially FA, AA, and IA, were frequently found to extend the guidelines of KMOE [13]. According to the toxic release inventory (TRI) data of the US EPA, acetaldehyde is also classified as a known or suspected carcinogen [40,41]. The reference compound, formaldehyde (>100 ppb) is also well known to cause irritation of eyes, nose, and throat, while being capable of accelerating asthma symptoms and other respiratory illnesses [42–44].

In addition, the primary acute effect of inhalation exposure to acetaldehyde includes irritation of the eyes, skin, and respiratory tract in humans, while at higher exposure levels, erythema, coughing, pulmonary edema, and necrosis may also occur [45]. In this



Fig. 2. Comparison of odorant concentrations between different countries of charcoal production (single letter symbol for country code).

investigation, we found several charcoals yielded a high concentration of ammonia which exceeded the guideline levels [13]. A recent study revealed that ammonia gas can cause damage, when hydrous ammonia reacts with tissue water to form strongly alkaline solution, ammonium hydroxide. It is a product from an exothermic reaction and can cause significant thermal injury to the skin, eyes, and especially to the respiratory system [46].

4. Conclusion

According to this study, several odorants (toluene, p-X, S, MEK, AA, PA, IA, and NH₃) are often significantly emitted during charcoal combustion and thereby play an important role in odor pollution under certain circumstances. Although emissions of many odorants from different origins were below harmful levels, a few odorants (AA, PA, IA, and NH₃) were found to exceed certain guidelines set for odorant emission control. If the strength of odor emission is compared in terms of sum of odor intensity (SOI), the values varied in the range of 2.51 to 5.66. As such, the emission levels of odorants from some charcoal products were significant enough to cause strong odor pollution. An evaluation of odorant levels released from charcoal combustion suggests the possibility that the materials used for charcoal production may play a vital role in terms of odor emission. Apart from odorants, our analysis confirmed that emission of other health hazardous reference components (benzene, CS₂, SO₂, CO, THC, and PM₁₀) can occur considerably from charcoal combustion. Given this situation, efforts should be made to properly establish regulation guidelines for charcoal production with respect to odor as well as other airborne pollutant emission. The results of a certain charcoal product like the one product made in the US suggest the possibility that odorant emission levels can be significantly reduced with proper treatment (during production). While seeking safer products for use, due to hazardous health effects especially in indoor environments, proper ventilation should also be implemented to reduce adverse health impacts from charcoal use.

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